

# Close Localization of the Genes for GM-CSF and IL3 in Human Genome

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Colony-stimulation factors (CSFs), a family of glycoprotein growth factors, have been shown to support clonal proliferation of hematopoietic progenitor cells in vitro [1]. Macrophage-CSF (M-CSF or CSF-1) [2] and granulocyte-CSF (G-CSF) [3] stimulate cells committed to the macrophage and granulocyte lineages respectively, whereas granulocyte-macrophage-CSF (GM-CSF) and interleukin-3 (IL3 or multi-CSF) are capable of stimulating proliferation and differentiation of progenitors along multiple pathways.

Successful cloning of cDNA and genomic copies of mouse and human genes for IL3 [3, 4] and GM-CSF [5, 6], as well as for M-CSF and G-CSF, have had a great impact on the analysis of biological properties of those molecules in vivo and in vitro [7, 8].

The GM-CSF and IL3 genes have been mapped to human chromosome 5 at bands q23-31 [9, 10], a region that is frequently deleted in patients with myeloid disorders [del(5q)] [11]. Several other growth factors and growth-factor receptors – in particular, the CSF-1 gene and proto-oncogene FMS, coding a protein possibly identical to the receptor for CSF-1 – are also located within this region of chromosome 5 [12]. There is a possibility that a family of genes responsible for regulation of cell growth during hematopoiesis is located within the limited segment of chromosome 5 [9]. Precise mapping of this region is essential to

the understanding of functional relationships between the genes and could reveal the genes for other growth factors and their receptors that may be located within this region.

For isolation of genomic DNA clones containing genes for human GM-CSF and IL3 we prepared from human leukocyte DNA a genomic library of  $1.5 \times 10^6$  clones. Using synthetic oligonucleotides, we identified eight individual clones that hybridized with the IL3 probe and five clones that hybridized with the GM-CSF probe; three of these hybridized with both probes. Southern blot hybridization analysis of DNA restriction fragments of individual phages revealed that three independently cloned 20-kb fragments of human genomic DNA contain sequences of both IL3 and GM-CSF genes. Physical maps of the isolated clones are shown in Fig. 1.

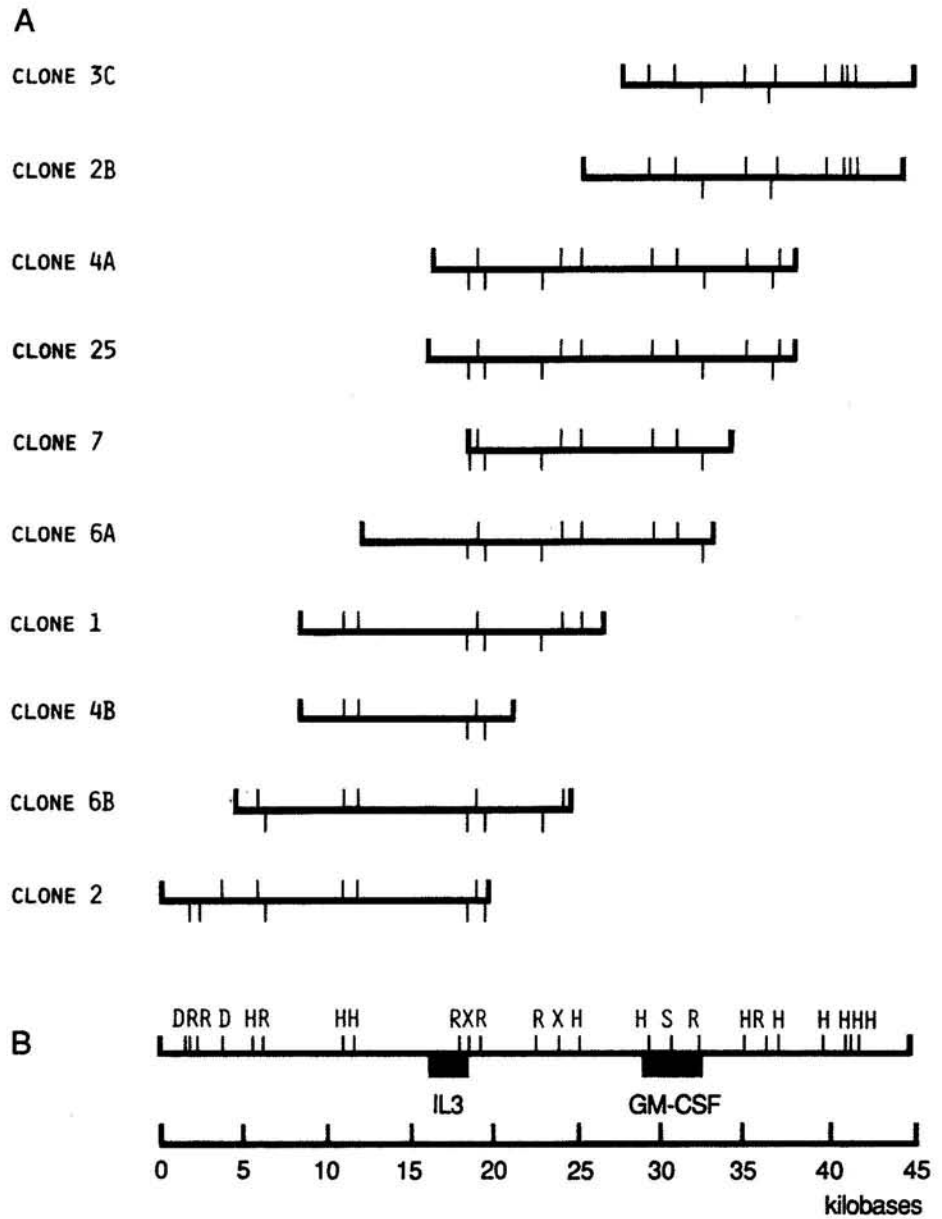
Next, we analyzed localization of the two genes in human placental DNA using Southern blot analysis (Fig. 2). The fragments generated by *Xba*I restriction endonuclease hybridized with both IL3 and GM-CSF probes.

The results strongly indicate a close genomic linkage of human IL3 and GM-CSF genes. The distance between the genes is 10 kb, and they are arranged in head-to-tail fashion, the gene for GM-CSF following the gene for IL3.

Close linkage of the two CSF genes may indicate either that they have coordinate regulation during T-lymphocyte gene expression, or that they have diverged from a common ancestral gene by duplication. The latter hypothesis is supported by the fact that both genes have similar exon-intron structures and com-

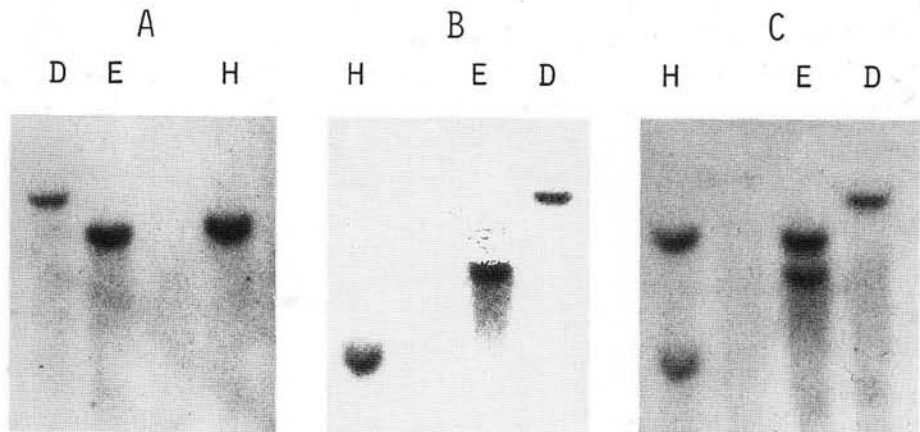
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**Fig. 1. A** Physical maps of the individual phage clones containing genes for human GM-CSF and IL3. The positions of the recognition sites for restriction endonucleases *EcoRI* (*R*), *HindIII* (*H*), *XhoI* (*X*), *Sall* (*S*), and *XbaI* (*D*) are indicated. **B** Physical map of the whole region of human genome containing the genes

**Fig. 2 A-C.** Southern blot analysis of human DNA. The DNA from human placenta was cleaved with *EcoRI* (*E*), *HindIII* (*H*), and *XbaI* (*D*) and hybridized with the probes for IL3 (**A**), GM-CSF (**B**), and with both probes together (**C**)



mon features in the secondary structure of the two polypeptides displayed in distribution of alpha-helical regions.

## References

1. Metcalf D (1984) The hematopoietic colony-stimulating factors. Elsevier, New York, Oxford, Chapter 11. Tissue and cellular sources of the colony stimulating factors, p 309–329
2. Ihle JN, Keller J, Oroszlan S, Henderson LE, Copeland TD, Fitch F, Prystowsky MB, Goldwasser E, Schrader JW, Palaszynski E, Dy M, Lebel B (1983) Biologic properties of homogeneous interleukin 3. I. Demonstration of WEHI-3 growth factor activity, mast cell growth factor activity, P cell-stimulating factor activity, colony-stimulating factor activity, and histamine-producing cell-stimulating factor activity. *J Immunol* 131:282–287
3. Cantrell MA, Anderson D, Cerretti DP, Price V, McKeregham K, Tushinski RJ, Mochizuki DY, Larsen A, Grabstein K, Gillis S, Cosman D (1985) Cloning, sequence, and expression of a human granulocyte/macrophage colony-stimulating factor. *PNAS* 82:6250–6254
4. Kaushansky K, O'Hara PJ, Berkner K, Segal GM, Hagen FS, Adamson JW (1986) Genomic cloning, characterization, and multilineage growth-promoting activity of human granulocyte-macrophage colony-stimulating factor. *PNAS* 83:3101–3105
5. Fung MC, Hapel AJ, Ymer S, Cohen DR, Johnson RM, Campbell HD, Young IG (1984) Molecular cloning of cDNA for murine interleukin-3. *PNAS* 81:233–237
6. Yang Yu-Ch, Ciarletta AB, Temple PA, Chung MP, Kovacic S, Witek-Giannotti JS, Leary AC, Kriz R, Donahue RE, Wong GG, Clark S (1986) Human IL-3 (Multi-CSF): identification by expression cloning of a novel hematopoietic growth factor related to murine IL-3. *Cell* 47:3–10
7. Donahue RE, Wang EA, Stone DK, Kamen R, Wong GG, Seghal PK, Nathan DG, Clark SC (1986) Stimulation of haematopoiesis in primates by continuous infusion of recombinant human GM-CSF. *Nature* 321:872–875
8. Kindler V, Thorens B, de Kossodo S, Allet B, Eliason JF, Thatcher D, Farber N, Vassalli P (1986) Stimulation of hematopoiesis in vivo by recombinant bacterial murine interleukin 3. *PNAS* 83:1001–1005
9. Pettenati MJ, LeBeau MM, Lemons RS, Shima EA, Kawasaki ES, Larson RA, Sherr CJ, Diaz MO, Rowley JD (1987) Assignment of CSF-1 to 5q33.1: Evidence for clustering of genes regulating hematopoiesis and for their involvement in the deletion of the long arm of chromosome 5 in myeloid disorders. *PNAS* 84:2970–2974
10. LeBeau MM, Epstein ND, O'Brien SJ, Nienhuis AW, Yu-Chung Yang, Clark SC, Rowley JD (1987) The interleukin 3 gene is located on human chromosome 5 and is deleted in myeloid leukemias with a deletion of 5q. *PNAS* 84:5913–5917
11. Mitelman F (1985) In: Sandberg A (ed) *Progress and topics in cytogenetics*, vol 5, p 107. Liss, New York
12. LeBeau MM, Westbrook CA, Diaz MO, Larson RA, Rowley JD, Gasson JC, Golde DW, Sherr CJ (1986) Evidence for the involvement of GM-CSF and FMS in the deletion (5q) in myeloid disorders. *Science* 231:984–987